Acute increase in plasma D-dimer level in ovarian torsion: an experimental study

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BACKGROUND: Torsion of the ovary is a rare but serious cause of gynecologic surgical emergency. Specific laboratory markers that support the preoperative diagnosis of ovarian torsion are not currently available in the clinical routine. The aim of this study was to investigate the diagnostic value of plasma D-dimer level as an early indicator of ovarian torsion in an experimental rat ovarian torsion model.

METHODS: Sixteen female adult Sprague–Dawley rats were used for this controlled experimental study. Eight rats in the sham operation group (Group I) underwent a surgical procedure similar to Group II but the ovary was not occluded. In Group II (eight rats), a torsion model was created by using atraumatic vascular clips just above and below the right ovary for a 2-h period of ischemia. Right ovaries were surgically removed at the end of the procedure in each group. Blood was sampled before and after operation to assess plasma D-dimer levels. The main outcome measure was ovarian histopathologic findings scores and plasma D-dimer levels.

RESULTS: There was no significant difference in pre-operative plasma D-dimer levels (0.5963 ± 0.2047 mg/l in Group I, 0.6344 ± 0.1348 mg/l in Group II, P = 0.815, Mann–Whitney U-test). However, mean plasma D-dimer value for Group II was significantly higher than that in the control group (1.2267 ± 0.3099 versus 0.6213 ± 0.2346 mg/l, respectively, Mann–Whitney U-test, P < 0.001), following 2 h of ovarian torsion. Ovarian tissue damage scores were also statistically significantly different among groups.

CONCLUSIONS: If the observations made in a rat model are extended to humans, plasma D-dimer measurement may be a valuable parameter in the early diagnosis of ovarian torsion.

Key words: biochemical marker / ovarian torsion / D-dimer / diagnostic parameter

Introduction

Torsion of the ovary is a rare but serious cause of gynecologic surgical emergency with a prevalence of 2.7% (Hibbard, 1985). Rapid diagnosis and surgical treatment are the keys to preserve fertility and salvage the twisted ovary. However, the clinical diagnosis of ovarian torsion usually relies only on non-specific clinical signs, which delay the diagnosis, such as the presence of abdominal pain and ultrasonographic finding of an adnexal mass. But a worrying fact is that most (56%) of the patients treated by an urgent diagnostic laparoscopy do not have a confirmed ovarian torsion (Cohen et al., 2001). Specific laboratory markers that support the preoperative diagnosis of ovarian torsion are not available in the clinical routine today. A statistically significant association between the diagnosis of ovarian torsion and interleukin-6 (IL-6) in peripheral blood was demonstrated previously (Cohen et al., 2001; Daponte et al., 2006). Additionally, in the rat ovarian torsion model, ischemia-modified albumin (IMA) was found to be associated with torsion response (Aran et al., 2010).

The use of D-dimer levels as an indicator of intestinal ischemia has gained wide popularity following the publication of a number of studies indicating the potential useful role of D-dimer levels in diagnosis of venous thromboembolic disorders in all body organs, including the lungs, upper extremities, pelvis, thigh and calf (Acosta et al., 2004). A similar pathophysiology could be expected in ovarian torsion in which there were firstly venous and lately arterial thrombotic vessels in the twisted pedicle and the ischemic ovary (Hibbard, 1985). To the best of our knowledge, there is no such study reporting the diagnostic value of D-dimer in ovarian torsion.
The aim of this experimental study was, therefore, to investigate the effect of ovarian torsion on plasma D-dimer levels and to determine whether D-dimer levels were a useful adjunct that could be used in the diagnosis of ovarian torsion.

Materials and Methods

Animals and study design
A total of 16 female adult Sprague–Dawley rats weighing 150–220 g were obtained from the Karadeniz Technical University’s Experimental Animal Laboratory. The animals were kept under standard laboratory conditions at a controlled temperature of 20–22°C, humidity of 55–60% and controlled photoperiod of 12:12 h light:dark. They were provided with a standard laboratory diet, free access to food and water, and were housed at the Animal Research Center of Karadeniz Technical University. The study protocol was reviewed and approved by Karadeniz Technical University Ethics Committee for animal research.

Randomization
To randomly allocate the animals into groups, all rats were initially randomly numbered. Then, computer-assisted randomization was utilized according to the instructions at www.randomization.com. The study team was blind to the randomized groups, at least until the randomization was carried out, while the investigators responsible for the biochemical and histological studies were all blind to the randomized groups until the end of the study. Rats were allocated randomly to Group I (control) or Group II (ovarian torsion model) on the day of the experiment.

Anesthesia and surgical technique
Each rat was weighed and anesthetized with intraperitoneal ketamine hydrochloride (50 mg/kg Ketalar; Eczacibasi, Istanbul, Turkey) and xylazine hydrochloride (10 mg/kg Rompun; Bayer, Leverkusen, Germany), which were repeated as it is necessary to maintain anesthesia throughout the experiments. They were allowed to breathe spontaneously (not intubated) during the study. The animals were oxygenated with an oxygen mask, through which flowed a volume of 1–1.5 l/min in the first part of the procedure and throughout the experiment, but oxygen saturation was not monitored.

Rats were placed in the dorsal recumbent position and covered with sterile drapes during surgery. A 1-ml preoperative blood sample was drawn from the tail of each rat to determine basal plasma levels of D-dimer. Then, the skin area of the incision was shaved and disinfected. A longitudinal incision of 2 cm was performed in the midline area of the lower abdomen for laparotomy, and the uterine horns and adnexa were located. In Group I (control group, sham-operated group), the abdominal wall was kept open for 1 min and then closed with 3/0 silk sutures. In Group II, a torsion model was created by using atraumatic vascular clips just above and below the right ovary. Incision was closed with 3/0 silk sutures. In both groups, 2 h after the first operation, re-laparotomy was performed through the previous incision sites. A 1-ml blood sample was taken again from the tail of each rat to determine the effect of the sham operation (Group I) and torsion operation (Group II) on plasma D-dimer level. After blood samples had been taken, the right ovary was surgically removed.

Histologic examination
The ovarian tissues were fixed in 10% neutral buffered formalin solution for 48 h, dehydrated, cleared and embedded in paraffin as usual. Serial tissue sections at a thickness of 5 μm were cut using the microtome and stained with hematoxylin and eosin (H&E) for general morphological observation. All sections were investigated with a light microscope (Olympus BX-51; Olympus Optical Co., Ltd., Tokyo, Japan) and the pieces were photographed. At least five microscopic areas were examined to semi quantitatively score the specimens. The criteria for ovarian injury were follicular cell degeneration, vascular congestion, hemorrhage and infiltration by inflammatory cells. Each specimen was scored for each criterion, using a scale ranging from 0 to 3 (0: none; 1: mild; 2: moderate; 3: severe) (Güven et al., 2010). The ovary sections were analyzed in a blinded fashion by the same histologist.

D-dimer measurement
Blood samples collected in citrated tubes were centrifuged for 15 min at 1000 rpm at room temperature. Plasma levels of D-dimer in separated blood samples had been taken, the right ovary was surgically removed. plasma levels of D-dimer in separated were quantified using an immuno-turbidimetric assay (STA Liatest D-dimer, Diagnostica Stago, Asnières-sur-Seine, France) on the STA-R coagulation analyzer. STA-Liatest D-dimer is a latex-based immunoassay obtained from Diagnostica Stago. The assay was performed using the STA-R equipment from Diagnostica Stago. The detection limit of this assay is 0.22 mg/l of D-dimer. STA-Liatest D-dimer assay was expressed in milligrams per liter of fibrinogen equivalent units, and the intra- and inter-assay coefficients of variation were 3.0 and 3.8%, respectively. The time required to have D-dimer result available for clinical decision-making is almost 40 min by using immuno-turbidimetric assay on the STA-R coagulation analyzer (manufacturer data).

Statistical analysis
The Statistical Package Program for the Social Sciences (SPSS 11.0; SPSS Inc., Chicago, IL, USA) was used for statistical analysis. Data normality were assessed with the Kolmogorov–Smirnov test. Statistical comparisons between groups were performed using the Mann–Whitney U (for unrelated samples) and Wilcoxon (for related samples) tests. The correlation coefficient was determined by the Pearson test. All P-values were two-tailed and statistical significance was set at P < 0.05.

Results

The mean body-weights of rats in Groups I (190.00 ± 12.25 g) and II (193.75 ± 28.25 g) were comparable (P = 0.743, Mann–Whitney U-test). There was no significant difference in pre-operative plasma D-dimer levels (0.5963 ± 0.2047 mg/l in Group I, 0.6344 ± 0.1348 mg/l in Group II, P = 0.815, Mann–Whitney U-test). However, mean plasma D-dimer value for Group II was significantly higher than that in the control group (1.2267 ± 0.3099 versus 0.6213 ± 0.2346 mg/l, respectively, Mann–Whitney U-test, P < 0.001), following 2 h of ovarian torsion.

A comparison of individual point for post-operative plasma D-dimer level in each group is given in Fig. 1. There was a significant increase in the plasma D-dimer value in Group II [P = 0.008, Wilcoxon test (including baseline values)], while no significant change was observed in Group I [P = 0.528, Wilcoxon test (including baseline values)] during the experiment. Compared with Group I (mean difference from the baseline +0.0250 ± 0.2660 mg/l), ovarian torsion significantly increased the plasma levels of D-dimer in Group II (mean difference from the baseline +0.5922 ± 0.3001 mg/l). The comparison was also statistically significant (P < 0.001, Mann–Whitney U-test).

A comparison of histopathological examination scores is shown in Fig. 2. Compared with Group I, in Group II, histologic specimens of the ovary had higher scores for follicular cell degeneration, vascular...
congestion, hemorrhage and inflammatory cell infiltration. As shown in Fig. 3a–c, compared with the control group, in the ovarian torsion group, severe follicular cell degeneration, vascular congestion, hemorrhage and inflammatory cell infiltration were observed. The mean histopathologic score (the summary score of four ovarian injury parameters) was significantly higher in Group II than in Group I (7.6 ± 2.2 versus 2.1 ± 1.0, respectively, Mann–Whitney U-test, P = 0.001).

Bivariate analysis revealed that post-operative plasma D-dimer level was well correlated with the summary score of four ovarian injury parameters (r = 0.633, P = 0.006).

**Discussion**

In this study, D-dimer, which reflects the extent of fibrin turnover, was studied as a possible marker for ovarian torsion. Our results demonstrate that plasma D-dimer levels appear to be elevated in ovarian torsion and may be useful in the early diagnosis and management of such surgical emergency conditions.

The diagnosis of ovarian torsion is usually done under urgent conditions of the gynecological emergency room. Moreover, several other gynecological and non-gynecological pathologies must be excluded. Urgent laparoscopy is considered the gold standard for accurate diagnosis. Thus many gynecologists elect to perform an urgent diagnostic laparoscopy rather than to delay the definite diagnostic procedure. But a worrying fact is that most (56%) of the patients treated by an urgent diagnostic laparoscopy do not have a confirmed ovarian torsion. The risk associated with urgent laparoscopy, often undertaken during the night, especially when performed by inexperienced personnel, may be increased. It is well known that under such urgent conditions, the rate of complications is increased (Cohen et al., 2001). A simple and more precise test in the sera may assist in making the diagnosis. If this test is found to be accurate, then patients may benefit from a justified urgent laparoscopy due to ovarian torsion rather than risk damage to the ovary following conservative observation. Additional imaging modalities may further delay the decision to operate and are not always helpful (Pena et al., 2000).

Serum IL-6, IL-8, tumor necrosis factor-alpha (TNF-a) and E-selectin have been investigated as serum markers in patients with ovarian torsion (Cohen et al., 2001; Daponte et al., 2006). A mouse model of myocardial ischemia reperfusion indicated that IL-6 was significantly elevated during the reperfusion phase (Hirano et al., 1990). A similar mechanism might exist in ovarian torsion, and it was previously reported, in eight patients, that a significant association between IL-6, but not TNF-a, and diagnosis of ovarian torsion (Cohen et al., 2001) exists. It has also been found that patients with serum IL-6 values > 10.2 pg/ml had a 16 times higher risk of having ovarian torsion (Daponte et al., 2006). Estimation of serum IL-6 level in a female patient with vague clinical signs of possible ovarian torsion such as adnexal mass and acute abdominal pain could help in differentiating patients, who would benefit from emergency surgical intervention, from others who could be treated conservatively. Among situations investigated and treated surgically, IL-6 was also associated with intestinal ischemia or endometrioma and appendicitis (Daponte et al., 2006). Additionally, in a recently published well-designed experimental rat ovarian torsion model study, elevated serum IMA level was found to be associated with torsion response (Aran et al., 2010).

A mouse model of intestinal ischemia reperfusion indicated that D-dimer was significantly elevated during the reperfusion phase. In acute intestinal ischemia, local intravascular coagulation is activated, thrombin formation and conversion of fibrinogen into fibrin increase, and intravascular and extravascular fibrin accumulates. As a result, the levels of thrombin, antithrombin complex and D-dimer might increase (Schoots et al., 2003). A similar mechanism might exist in ovarian torsion. A spectrum of ischemic changes due to ovarian torsion is noted in the ovary. The amount of damage depends on the extent of vascular compromise. First, the venous flow and lymphatic return are blocked, along with continued arterial perfusion of the ovary, leading to diffuse enlargement and edema of the ovarian parenchyma and often to peripheral follicular distension due to transudation of the fluid into the cysts. With further passage of time, continued edema and increased pressure on the twisted pedicle will cause venous
occlusion followed by arterial thrombosis. Thus, we preferred to study a rat model to show elevated D-dimer levels in ovarian torsion.

Clinical studies of ovarian torsion have shown that early diagnosis is the key to a successful outcome (Cohen et al., 2001; Daponte et al., 2004; Aran et al., 2010). Therefore, any new marker should have significantly increased concentrations immediately following the initiation of the clinical pathology that results in ovarian torsion. D-dimer can be an appropriate marker in this sense because it is the most rapidly elevating marker among fibrinolytic products: in bleeding esophageal varices, it increases significantly even 5 min after endoscopic embolization with thrombin (Suehiro and Koyama, 1991). The same increase can be anticipated after ovarian torsion, and this may aid in rapid diagnosis and treatment of ovarian torsion. We measured plasma D-dimer levels 2 h after the operation because various studies have shown that D-dimer increases within minutes after the onset of ischemia and remains elevated for 6–12 h (Acosta et al., 2004; Kurt et al., 2005; Altinyollar et al., 2006).

The limitation of D-dimer testing in surgical practice is its low specificity in many conditions. Every surgical trauma and tissue injury or even i.m injection itself can activate coagulation and consequently the fibrinolytic system and cause elevated D-dimer values (Acosta et al., 2004). In the present study, control group (sham-operated group) was designed in order to interpret the D-dimer test results in rats with torsed ovary. Thus, it was expected that mean D-dimer levels in both sham and torsion operation groups in the second post-operative hour were higher than that in the preoperative. But second post-operative hour mean D-dimer plasma levels in rats with torsed ovary were found statistically significantly higher than that in the pre-operative and second post-operative hour mean D-dimer plasma levels in sham operation group.

This study also had some limitations in terms of the model employed. This work was a case-controlled study, but it may not mimic typical ovarian torsion cases seen in clinical practice. However, the rarity of ovarian torsion and its subtle symptoms make a wider series of clinical studies difficult. The absence of serial blood measurement in this experimental model may be the limitation of our study. Serial blood measurements could alter rat hemodynamic status and affect plasma D-dimer levels; thus, we preferred not to use serial measurements.

The results obtained from the current experimental study suggest that the measurements of D-dimer might be used as a plasma marker in early detection and diagnosis of ovarian torsion. However, further clinical and experimental studies of a larger size are required to clearly confirm that an elevated D-dimer level is a useful adjunct in the early diagnosis of ovarian torsion.

**Authors’ roles**

The study was designed by C.K., T.A. and S.G. Data were collected by C.K., T.A., S.G.K. and E.Y. Data were analyzed by C.K., T.A. and S.G., who also wrote the first draft of the manuscript. All authors participated in the finalization of the manuscript and read the final draft.

**Conflict of interest:** none declared.

**References**


